Conserved Ribosomal Protein Sequences S5, S18, S27, S30 in the Pacific Oyster Crassostrea gigas and Crassostrea virginica

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Abstract

cDNA fragments encoding ribosomal proteins were isolated from the mantle of pacific oyster *Crassostrea gigas* by the subtractive hybridization method. The sequence information was used to isolate entire cDNAs, and the predicted amino acid sequences were shown to be very similar to the 40S ribosomal protein genes (S5, S18, S27, and S30) of *Crassostrea virginica*. Northern blot hybridization revealed that S5, S18, and S30 were predominantly expressed in the digestive gland, the gill, and the mantle. On the other hand, S27 was highly expressed in the adductor muscle.

1. Introduction

The mantle surrounds the whole body of a mollusc and is responsible for shell formation. Its important role in shell formation is to produce organic matrix proteins that are secreted into the extrapallial space and then regulate the crystallization of calcium carbonate. The process of shell formation, calcification, begins with the determination of which type of crystal, aragonite or calcite, should be made. Some organic matrix proteins have been isolated⁽¹⁻¹⁰⁾ and shown to regulate the formation of the two types of calcium carbonate crystal ^(11,12). These studies indicate that various organic matrix proteins have been found and have evolved in molluscs. We have been undertaking, by use of suppression subtractive hybridization (SSH) method, a study to identify the genes expressed in the mantle of the pacific oyster *Crassostrea gigas*⁽¹³⁾. In the analyzed sequences, 4 ribosomal proteins (S5, S18, S27, S30) were identified. Ribosomes consist of many different proteins, of which sequences are reported mainly in vertebrates and several model species. In contrast, comparatively little is known about ribosomal protein S27, which has a zinc finger domain, is expressed abundantly in the adductor muscle.

2. Results and Discussion

Because the 4 cDNAs encoding ribosomal protein were partial, to determine the whole nucleotide sequences, we isolated the 5'-end and 3'-end cDNAs by the RACE method using oligonucleotides specific to the 4 cDNAs as primers. The nucleotide sequence of the ribosomal protein S5 showed an open reading frame consisting of 612 bp followed by a canonical polyadenylation signal and poly (A) tail (Fig. 1). The predicted amino acid sequence showed 99% identity to the *C. virginica* S5 sequence (Fig. 5). *C. gigas* S5 was nearly equal in identity to human and mouse S5 (91% and 90% respectively), while it showed 84% identity to *Drosophila* S5, which is slightly less than the identity between the human and *Drosophila* sequences. The sequenced cDNA of S18 is 523bp in length and showed a 456 nucleotide open reading frame coding for a 152 amino acid sequence (Fig. 2). The predicted protein showed 99% identity to *C. virginica* S18 (Fig. 5) and 83% identity to human and mouse S18. To *Drosophila* S18, it showed 78% identity to *C. virginica* S18 (Fig. 5) and 93% identity to human and *Drosophila* sequences. The sequenced cDNA of S27 contained a single ATG-initiated open reading frame of 252bp (Fig. 3). The predicted protein showed 100% identity to *C. virginica* S18 (Fig. 5) and 93% identity to human and mouse S27. To *Drosophila* S27, it showed 80% identity, which is slightly less than the identity between the human and *Drosophila* sequences. The sequenced cDNA of S30 showed an open reading frame that starts at position 39 and includes 131 amino acids (Fig. 4). In several eukaryotic organisms, ribosomal protein S30 was shown to be transcribed as a single mRNA that is a fusion

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with a ubiquitin-like sequence⁽¹⁴⁾. As a result, a fusion protein is translated, then undergoes posttranslational processing to yield the S30 protein and the ubiquitin-like protein, respectively. Similarly the *C. gigas* S30 cDNA showed a fusion sequence composed of the ribosomal protein and the ubiquitin-like protein. When compared to *C. virginica* S30, the amino-terminal ubiquitin-like protein region showed 90% identity and the calboxy-terminal ribosomal protein S30 region showed 98% identity (Fig. 5). This difference of the identities indicates that two regions (ubiquitin-like protein and ribosomal protein S30) of the fusion gene have evolved under different selective pressure. Comparison of the determined sequence of 4 ribosomal proteins of *C. gigas* with counterparts of *C. virginica* showed striking identity, indicating that *C. gigas* and *C. virginica* are highly correlated evolutionarily. This notion is consistent with the result obtained from sequence analyses of 18S rRNA and 28S rRNA in many mollusc species⁽¹⁵⁾.

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- 1 TGTTAACGGCACTATGACTGAGAACTGGGATGAGCCTGCTCCGGCAGTAGAATTGCCAGA M T E N W D E P A P A V E L P E 16
- 61 AATCAAGCTCTTTGGCAAATGGTCATCAGATGATGATGTCCAAGTCAGCGACATCAGTTTAAC I K L F G K W S S D D V Q V S D I S L T
- 121 TGATTACATTGCTGTCAAAGAGAAGTATGCAAAATATTTGCCACACTCCTCAGGCAGATA D Y I A V K E K Y A K Y L P H S S G R Y
- 181 CCAAGTAAAGAGATTTAGAAAATCACAGTGCCCAATTGTTGAACGCCTGACATGTTCACT Q V K R F R K S Q C P I V E R L T C S L
- 241 TATGATGCATGGAAGAAACAATGGAAAGAAACTCTTGACAACCCGCATTGTGAAACATGC M M H G R N N G K K L L T T R I V K H A 96
- 301 CTTTGAAATCATTCACTTGCTCACAGGAGAAAACCCCTCTCCAAGTTTTGGTGAATGCCAT F E I I H L L T G E N P L Q V L V N A I 116
- 361 CATCAACAGTGGCCCCCGTGAGGACTCCACTCGTATTGGTCGTGCTGGTACCGTCAGGCG I N S G P R E D S T R I G R A G T V R R 136
- 421 TCAGGCTGTGGACGTCTCCCACTGAGGCGTGTCAACCAGGCCATCTGGCTCCTGTGTAC Q A V D V S P L R R V N Q A I W L L C T
- 481 CGGGGCACGTGAAGCCTCCTTCAGGAATATCAAGACTATTGCTGAGTGTTTGGCTGATGA G A R E A S F R N I K T I A E C L A D E 176
- 541 GCTGATCÅATGCTGCCAAGGGATCTTCAAACTCCCATGCCATCAAGAAGAAGGATGAATT L I N A A K G S S N S H A I K K K D E L
- 601 GGAACGTGTTGCCAAGTCCAACCGATAAACTATTTACTGTGCTTTCTGTGAACAGGAAAA E R V A K S N R *
- 661 TAAACTGTCAGGCA
- 1 CTTTTCCGCTGAATTATCAACAATGGCTTTGATACTGCCAGAGAAGTTTCAGCACATTCT M A L I L P E K F Q H I L
- 61 TCGTATCCTCAACACAAATATTGATGGACGAAGGAAAATTATGTTCGCTATGACTGCCAT R I L N T N I D G R R K I M F A M T A I
- 121 CAAGGGTATCGGTAGACGATATGCTAATGTTGTCTGCAAGAAAGCTGATGTAGATATCAC K G I G R R Y A N V V C K K A D V D I T
- 181 AAAAAGGGCAGGGGAACTCTCAGAAGAAGAGATTGACAAAATTGTCACAAATTATGCAGAA K R A G E L S E E E I D K I V T I M Q N
- 241 CCCTCGTCAGTACAAGATTCCTGACTGGTTCCTTAACAGGCAGAAGGACATTAAGGATGG P R Q Y K I P D W F L N R Q K D I K D G
- 301
 TAAATTCAGCCAGGTCATGTCCAACACACACGGACAACAAACTCCGTGAGGATCTGGAGCG

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- 361 ACTAAAGAAGATCCGAGCACACAGAGGTCTCCGTCACTACTGGGGTCTAAGAGTGAGAGG L K K I R A H R G L R H Y W G L R V R G 133
- 421 TCAGCACACAAAGACCACAGGAAGAAGAAGAAGAAGCTGTTGGTGTGGGCCAAGAAGAAGTA Q H T K T T G R R G R T V G V A K K K * 152
- 481 AACTGTGTAAATGAGGGAGGAAAAGTAAAAAAAGTGTATCCA

Fig. 1 Nucleotide and deduced amino acid sequence of ribosomal protein S5.

The nucleotide positions are indicated on the left and the amino acid positions are indicated on the right. The putative polyadenylation sequence is underlined. The stop codon is marked with an asterisk. DDBJ accession number: AB199894

Fig. 2 Nucleotide and deduced amino acid sequence of ribosomal protein S18.

The nucleotide positions are indicated on the left and the amino acid positions are indicated on the right. The stop codon is marked with an asterisk. DDBJ accession number: AB199895

To investigate the expression of 4 ribosomal proteins of C. gigas, Northern blot analysis was conducted using cDNA fragments as probes. The signals of 4 transcripts were stronger in the mantle than in the egg. This finding is in agreement with the fact that the cDNAs were isolated as sequences expressed highly in the mantle by the SSH method. In invertebrates, early development is relatively independent of newly synthesized transcripts, and zygotic gene activation is usually observed after the cleavage stage. Low expression of the 4 ribosomal protein genes suggests that translational activity is relatively low in the eggs of molluscs. As for S5, S18, and S30, hybridization signals were equally detected in the digestive gland, the gill, and the mantle. S27 was expressed highly in the adductor muscle. It is reported that the ribosomal protein S27 is differentially expressed in the tissues of mammals⁽¹⁶⁾. In human periodontal ligament cells, which connect the teeth and the dental alveoli, S27 expression is induced by mechanical stress⁽¹⁷⁾. Both the periodontal ligament in mammals and the adductor muscle in molluscs are fibrous tissue and are exposed to mechanical stress, suggesting that the ribosomal protein S27 is involved in the morphogenesis and maintenance of these tissues in both vertebrates and invertebrates.

- ATGCCTCTCGCTAAAGATTTATTGCATCCCTCTTTGGAGGAGGAGAAAAGAACATGCAAA 1 M P L A K D L L H P S L E E E K R T C K
- 61 CTGAAGAGATTGGTCCAAAGTCCAAACAGTTATTTCATGGATGTTAAATGTCCAGGATGC L K R L V Q S P N S Y F M D V K C P G C
- TACAAGATTACCACAGTTTTCAGCCACGCCCAGACGGTGGTGTTATGTGTGGGGGTGCTCC 121 YKITTVFSHAQTVVLCVGCS
- 181 ACAGTGCTGTGCCAGCCCACCGGGGGGAAAGCCAGACTCACAGAGGGCTGCTCCTTCCGT T V L C O P T G G K A R L T E G C S F R
- 741 AAGAAGTCGCACTAGACTGACAACGTTATTTATATAGGCTGTATGAGAAAGAGAGGGCTGA K K S H *
- 301 GATATTTGGGGTTGGCATGGCGACCGTCGTCAACATTTGGACTTTGTGATACTCTTCATT
- TATTTTTTTTTTGGACAACACGCATAATTGTTTCTTTGATTTACGACCCTGGGTCAGA 361
- 421 TCATCAATCGTCTCAGAAGATTCCACACCCAGGCTTTG
- 1 MQLFVRGS
- GCGAGACTCATGCCTTGCAGTTGGCAGGAAACGAAACTGTGTCCGACATCAAGAATTTGA 61 THALQLAGNE TVSDIKNL 28 I
- TTAGCAAGAGAGAGGGGTTTCCAGTTGAGGAACAGATTATTCTGTATGCCGGCAAACCAC 121 SKREGFPVEEQIILYAGKP 48
- 181 TTCAGGATGAATATGAGTTGACCAAACTTAATGACTTGTCCACCCTGGACATTGAAGTCA QDEYEL TKLNDLSTLDIE V R 68
- 241 GAATGCTTGGAGGTAAAGTCCATGGCTCTCTTGCTCGTGCCGGAAAAGTCAAGGGACAGA M L G G K V H G S L A R A G K V K G Q T 88
- 301 CCCCAAAGGTTGAGAAACAAGAGAAGAAGAAGAAGAGGGGACAGGAAGGGCCAAGAGACGCA 108 P K V E K Q E K K K Q R T G R A K R R M
- TGCAGTACAACAGAAGATTTGGAGTTGTCGTCTCTACATTCGGTCGCAGAAAGGGACCCA 361 128 O Y N R F G V V V S T F G R R K G P Ν
- 421 ATGCTAACTCCTAAAATGTGTTTCTATCAAAAACTGGATTAAAATTTCGTTGTAGCTGCA 131 AN S

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- Fig. 3 Nucleotide and deduced amino acid sequence of ribosomal protein S27.
- 100 The nucleotide positions are indicated on the left and the amino acid positions are indicated on the right. The stop codon is marked with an asterisk. DDBJ accession number: AB199896

Fig. 4 Nucleotide and deduced amino acid sequence of ribosomal protein S30.

The ubiquitin-like protein region is boxed. The nucleotide positions are indicated on the left and the amino acid positions are indicated on the right. The stop codon is marked with an asterisk. DDBJ accession number: AB199897



Fig. 5 Northern blot analysis of 4 ribosomal proteins. (A) S5. (B) S18. (C) S27. (D) S30. (E) The same samples stained with ethidium bromide. Lane 1: digestive gland; Lane 2: gill; Lane 3: adductor

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4. References

- (1) Miyamoto, H., Miyashita, T., Okushima, M., Nakano, S., Morita, T., Matsushiro, A. (1996) A carbonic anhydrase from the nacreous layer in oyster pearls. Proc. Natl. Acad. Sci. USA 93, 9657-9660.
- (2) Sudo, S., Fujikawa, T., Nagakura, T., Ohkubo, T., Sakaguchi, K., Tanaka, M., Nakashima, K. (1997) Structures of mollusc shell framework proteins. Nature 387, 563-564.
- (3) Shen, X., Belcher, A. M., Hansma, P. K., Stucky, G. D., Morse, D. E. (1997) Molecular cloning and characterization of lustrin A, a matrix protein from shell and pearl nacre of Haliotis rufescens. J. Biol. Chem. 272, 32472-32481.
- (4) Samata, T., Hayashi, N., Kono, M., Hasegawa, K., Horita, C., Akera, S. (1999) A new matrix protein family related to the nacreous layer formation of *Pinctada fucata*, FEBS Lett. 462, 225-229.
- (5) Kono, M., Hayashi, N., and Samata, T. (2000) Molecular mechanism of the nacreous layer formation in Pinctada maxima. Biochem. Biophys. Res. Commun. 269, 213-218.
- (6) Weiss, I. M., Kaufmann, S., Mann, K., Fritz, M. (2000) Purification and characterization of perlucin and perlustrin, two new proteins from the shell of the molluse Haliotis laevigata. Biochem. Biophys. Res. Commun. 267, 17-21.
- (7) Marin, F., Corstjens, P., de Gaulejac, B., de Vrind-de Jong, E., Westbroek, P. (2000) Muncins and molluscan calcification. J. Biol. Chem. 275, 20667-20675.
- (8) Sarashina, I., Endo, K. (2001) The complete primary structure of molluscan shell protein 1 (msp-1), an acidic glycoprotein in the shell matrix of the scallop Patinopecten yessoensis. Mar. Biotechnol. 3, 362-369.

- (9) Zhang, Y., Xie, L., Meng, Q., Jiang, T., Pu, R., Chen, L., Zhang, R. (2003) A novel matrix protein participating in the nacre framework formation of pearl oyster, *Pinctada fucata*. J. Comp. Physiol B 135, 565-573.
- (10) Tsukamoto, D., Sarashina, I., and Endo, K. (2004) Structure and expression of an unusually acidic matrix protein of pearl oyster shells. Biochem. Biophys. Res. Commun. 320, 1175-1180.
- (11) Falini, G., Albeck, S., Weiner, S., and Addadi, L. (1996) Control of aragonite or calcite polymorphism by mollusk shell macromolecules. Science 271, 67-69.
- (12) Belcher, A. M., Wu, X. H., Christensen, R. J., Hansma, P. K., Stucky, G. D., and Morse, D. E. (1996) Control of crystal phase switching and orientation by soluble mollusc-shell proteins. Nature 381, 56-58.
- (13) Miyamoto, H., Hamaguchi, M., Okoshi, K. (2002) Analysis of genes expressed in the mantle of oyster *Crassostrea gigas*. Fish. Sci. 68, 651-568.
- (14) Olvera, J., Wool, I. G. (1993) The carboxyl extension of a ubiquitin-like proteins is rat ribosomal protein S30. J. Biol. Chem. 268, 17967-17974.
- (15) Giribet, G., and Distel, D. L. (2003) Bivalve phylogeny and molecular data, *In: Lydeard, C., Lindberg, D. R. (eds)* Molecular Systematics and Phylogeography of Mollusks. Smithonian Inst., Washington and London, pp. 45-90.
- (16) Thomas, E. A., Alvarez, C. E., Sutcliffe, J. G. (2000) Evolutionarily distinct classes of S27 ribosomal proteins with differential mRNA expression in rat hypothalamus. J. Neurochem. 74, 2259-2267.
- (17) Myokai, F., Oyama, M., Nishimura, F., Ohira, T., Yamamoto, T., Arai, H., Takashiba, S., Murayama, Y. (2003) Unique genes induced by mechanical stress in periodontal ligament cells. J. Periodont Res. 38, 255-261.

和文抄録

マガキ Crassostrea gigas と Crassostrea virginica において高度に保存された リボソームタンパク質 S5, S18, S27, S30

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マガキ Crassostrea gigas の 40S リボソームタンパク質 S5, S18, S27, S30 に関して、cDNA を単離し塩 基配列を決定した。予想されるアミノ酸配列は、いずれも Crassostrea virginica 種において明らかにさ れている配列と高い相同性を示した。S30 は、ユビキチン様ドメインと融合遺伝子として転写されており、 Crassostrea gigas と Crassostrea virginica においてひとつの転写ユニットであるにもかかわらず、ユ ビキチン様ドメインと S30 領域では、相同性において明らかな違いがみられ、モザイク的な進化が生じた 考えられる。発現様式は、それぞれのサブユニットごとに違いがみられ、S27 では、閉殻筋での強い発現 が認められた。